

**Appearance of systemic granulomatosis is modulated by the dietary
supplementation of vitamin E and C in meagre (*Argyrosomus regius*)
larvae fed inert microdiets**

Running title: Supplementation of vitamin E and C prevent granulomatosis in meagre larvae.

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Accepted refereed manuscript of:

Ruiz MA, Hernandez-Ruiz CM, Caballero MJ, Fernandez-Palacios H, Saleh R, Izquierdo MS & Betancor MB (2019) Appearance of systemic granulomatosis is modulated by the dietary supplementation of vitamin E and C in meagre (*Argyrosomus regius*) larvae fed inert microdiets [Supplementation of vitamin E and C prevent granulomatosis in meagre larvae]. *Aquaculture*, 506, pp. 139-147.

DOI: <https://doi.org/10.1016/j.aquaculture.2019.03.032>

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Abstract

Systemic granulomatosis has already been reported in meagre larvae with an adequate feeding protocol and enrichment media preventing its appearance in the first weeks of life. Afterwards, the control of this disease could be prevented through nutritional components of the inert food, being the antioxidants the key to success. For this reason, in the present study, meagre larvae were reared from 30 days post hatching (dph) with five isonitrogenous and isolipidic experimental microdiets with different levels of vitamin E and C: C- (40 mg kg⁻¹ E, 100 mg kg⁻¹ C), C+ (400 mg kg⁻¹ E, 1,000 mg kg⁻¹ C), Krill (400 mg kg⁻¹ E, 1,000 mg kg⁻¹ C and substitution of fish oil by krill oil), EC (200 mg kg⁻¹ E, 500 mg kg⁻¹ C) and EECC (800 mg kg⁻¹ E, 2,000 mg kg⁻¹ C). Prior to this, larvae were co-fed with rotifers and *Artemia* following a protocol which prevented the appearance of granulomas, as previously demonstrated. The substitution of fish oil by krill oil significantly increased levels of eicosapentaenoic acid (EPA, 16.6 %) and docosahexaenoic acid (DHA, 17.6 %) in meagre, consequently increasing the peroxidation index, which in turn translated into a higher incidence of granulomas. Although even low levels of vitamin E and C (40 mg kg⁻¹ E, 100 mg kg⁻¹ C; C-) allowed the adequate growth of larvae, these levels were not enough to prevent the appearance of granulomas, requiring superior levels of both antioxidant vitamins (800 mg kg⁻¹ E and 2,000 mg kg⁻¹ C) to mitigate systemic granulomatosis. This mitigation was simultaneous with the reduction of thiobarbituric acid reactive substances TBARs content in larvae, which were highly correlated with the appearance of granulomas ($R^2=0.892$, $y=0.0446x+0.0756$). A strong negative correlation was observed between the dietary levels of vitamin E ($y = -0.0098x + 11.174$, $R^2 = 0.8766$, $p \text{ value} = 0.019$, $r = -0.93$) and vitamin C ($y = -0.0022x + 6.4777$, $R^2 = 0.9278$, $p \text{ value} = 0.003$, $r = -0.96$) and the percentage of larvae with granulomas. The results showed that the occurrence of systemic granulomatosis seems to be associated to the larvae peroxidation status, so that high dietary levels of vitamin E and C (800 and 2,000 mg kg⁻¹, respectively; Diet EECC), reduced lipid peroxidation and completely prevented the appearance of granulomas in meagre larvae at 44 dph.

Keywords: meagre larvae, antioxidant vitamins, granulomatosis

1. Introduction

The whole life cycle of meagre (*Argyrosomus regius*) has been successfully closed, however there are still some challenges in meagre farming, being one of the more predominant ones the systemic granulomatosis. Systemic granulomatosis is a disease of unknown aetiology, although it has recently been evidenced that nutritional imbalances can promote its appearance (Ruiz et al., 2018a; Cotou et al., 2016). It is a non-infectious disease that affects internal organs, mainly liver, kidney and heart, where granulomas composed by a necrotic centre and surrounded by a layer of epithelial cells and macrophages are observed in the final stages (Ruiz et al., 2018a). It must be noted that the prevalence of systemic granulomatosis is so high in adult meagre that it can affect almost 100 % of population (Ghittino et al., 2004), being this stage too late to try to avoid the appearance of the disease. Nevertheless, granulomas have not only been detected in adult fish, but meagre larvae have also been found to show this histological alteration at very early stages (Ruiz et al., 2018b). In the afore mentioned study, granulomas were first described in liver and kidney at 20 days post hatching (dph) albeit differences were found among larvae fed the different dietary treatments/feeding sequences. In this sense, a co-feeding with rotifers (*Brachionus plicatilis*) and *Artemia* prior to weaning on an inert commercial microdiet proved to prevent the appearance of granulomas. On the other hand, when *Artemia* was not included in the feeding sequence, granulomas were detected from 20 dph although the incidence varied depending on the enrichment media used what again strengthens the hypothesis of a nutritional origin of the pathology. Therefore, a balanced nutrition during the first life stages of meagre could potentially prevent the development of systemic granulomatosis.

Imbalances in vitamins, particularly antioxidant vitamins such as vitamin E and C, have long been speculated to play a pivotal role in the appearance of systemic granulomatosis. Appearance of granulomas in gilthead sea bream (*Sparus aurata*) and turbot (*Scophthalmus maximus*) has been associated to a dietary deficiency of vitamin C (Paperna et al., 1980; Baudin-Laurencin et al., 1989; Coustans et al., 1990; Alexis et al., 1997). A deficiency in this nutrient causes an impairment of tyrosine catabolism, which leads to its precipitation in tissues and thereby the development of the granulomas

(Goldsmith, 1978). In previous studies, the combination of high dietary content of antioxidant vitamin E, C and K (15, 450 and 230 mg kg⁻¹, respectively) reduced the incidence of granulomas in juvenile meagre (Ruiz et al., 2018a). However, a high prevalence of granulomas was observed at the beginning of the experimental trial what prompted to evaluate the combination of vitamins at earlier life stages. If vitamins are to be blamed for the appearance of systemic granulomatosis, meagre larvae might be then at a higher risk of suffering the pathology as their higher growth and metabolic rates mean that vitamin requirements might be higher for larvae than juveniles or adult fish (Dabrowski, 1992). Additionally, limited information is available about the requirements of vitamin E and C in meagre larvae almost of the studies have been mainly focused on adults or juvenile fish. A recent study by El Kertaoui et al. (2017) showed that high levels of both vitamin E and C (1,500 and 1,800 mg kg⁻¹, respectively) improved growth and protection against oxidative stress in meagre larvae, but the effect of these antioxidant vitamins on the appearance of granulomas was not evaluated. Recently, the appearance of systemic granulomatosis has been observed to be affected by the fatty acid profile of the diet in meagre larvae, where the lowest supplementation of n-3 LC-PUFA (0.8 %) lead to a higher incidence of granulomas in liver (Carvalho et al., 2018). Docosahexaenoic acid (DHA, 22:6n-3) and eicosapentaenoic acid (EPA, 20:5n-3) are considered essential fatty acids in marine fish and are involved in the maintenance of structural and functional integrity of cell membranes (Izquierdo and Koven, 2011), normal growth (Rodríguez *et al.*, 1994; Salhi et al., 1997) and immune-deficiency (Izquierdo, 1996). The different fatty acid profile between fish oil and krill oil could have an impact on the appearance of granulomas, moreover, an absence of adequate levels of antioxidants, may lead to lipid oxidation as long as PUFA are available for oxidation (Hamre, 2011). Krill oil is higher in some fatty acids, such as EPA and DHA, compared to fish oil (Tou et al., 2007). Moreover, the phospholipid composition is different in both oils, in fish oil fatty acids are mainly stored as triglycerides, whereas in krill 30–65 % of the fatty acids are incorporated into phospholipids (Tou et al., 2007), which have higher bioavailability and are involved in regulation of more metabolic pathways (Ulven and Holven, 2015).

The overall aim of the present study was to evaluate the role of vitamin E and C in the appearance of systemic granulomatosis in weaned larvae (30 dph). Prior to the start of the study larvae were co-fed with rotifer and *Artemia* enriched with Easy DHA Selco as larvae fed this dietary regime did not show any granulomas at 30 dph in a previous trial (Ruiz et al., 2018b). Following this feeding sequence meagre will be fed four microdiets formulated to contain graded levels of inclusion of vitamin E and C. Additionally a fifth diet was formulated to contain krill oil as the single lipid source. Fish larvae growth and survival, histopathological evaluation and biochemical analysis were determined.

2. Materials and methods

2.1. Fish

Meagre eggs were obtained from an induced spawning from broodstock from the ECOAQUA facilities at University of Las Palmas de Gran Canaria (ULPGC; Telde, Canary Islands, Spain) where the experiment was carried out.

Rotifers were cultured at a density of 400 rotifers mL⁻¹ in 500 L enrichment troncoconical-tanks, with 80 % seawater and 20 % freshwater. Rotifers were enriched with Easy DHA Selco (INVE, Dendermonde, Belgium) (0.6 g L⁻¹) for 24 h. Meagre larvae were fed with enriched rotifers twice daily from 3 to 21 dph, before each feeding, rotifers were counted and added to maintained at a density of 10 rotifers L⁻¹ in the experimental tanks. *Artemia* cyst were hatched at 27 °C and 0.030 mg L⁻¹ salinity until 100 % hatch was achieved. Then, they were rinsed with seawater and transferred to a culture tank at 24 °C. *Artemia* was enriched with Easy DHA Selco (0.6 g million *Artemia*⁻¹) for 24 h before being fed to the larvae. Meagre larvae were fed with enriched *Artemia* from 12 to 30 dph following the protocol established by Ruiz et al., (2018b). Before each feeding, *Artemia* were counted and added to maintained at a density of 1.3-1.5 *Artemia* L⁻¹ in the experimental tanks. From 20 to 30 dph larvae were co-fed with *Artemia* and microdiet and fed microdiet only from 30 to 44 dph.

Larvae of 30 dph (total length 8.83± 0.65 mm, dry body weight 1.1 ± 0.01 mg) were randomly distributed in light grey colour cylindrical fibreglass experimental tanks (15 tanks; triplicate treatment) of 170 L capacity at a density of 3000 larvae tank⁻¹ and fed one of the five experimental diets for 14 days. All tanks were equipped with continuous

aeration and supplied with filtered UV-sterilized seawater at an increasing rate from 35% h⁻¹ to a 100% h⁻¹, to guarantee good water quality during the trial. Water entered the tanks at the bottom and exited at the surface. Oxygen (4.5-6.5 g L⁻¹), salinity (34 g L⁻¹) salinity and temperature (21.8 to 22.3° C) was daily measured. Photoperiod was kept at 12 h light: 12 h dark by fluorescent lights.

All procedures were conducted in accordance with the regulations set forward by the Spanish RD 53/2013 (BOE 8th February 2013) and the Directive 2010/63/EU of the European Parliament and of the Council of 22 September 2010 on the protection of animals used for scientific purposes. The experiment was subjected to ethical review by the Animal Welfare and Bioethical Committee at the University of Las Palmas de Gran Canaria (Ref 06/2018 OEBA ULPGC).

2.2. Diets

Five isonitrogenous and isolipidic experimental microdiets (pellet size 120-250 & 250-500 µm) were formulated (Tables 1 and 2). Krill meal was the source of protein whereas fish oil was the source of lipid, excepting for the diet labelled “Krill” in which krill oil was the single lipid source. Prior to preparing the feeds, the krill meal was defatted (three consecutive times with a chloroform: krill meal ratio of 3:1) to allow a better control of the fatty acid profile of the microdiet. A positive control diet (C+) was formulated based on the vitamin E (400 mg kg⁻¹) and C (1,000 mg kg⁻¹) levels found in a commercial microdiet (Gemma Micro 150 and 300 µm; Skretting, France). Based on this level of vitamins, other three diets with higher and lower levels of vitamin E and C was formulated, diet C-, Krill, EC and EECC (40/100, 400/1,000, 200/500 and 800/2,000 mg kg⁻¹ vitamin E and C respectively). Krill oil diet was formulated using krill oil as the only lipid source. Soy lecithin was used as a source of phospholipids, excepting in diet “krill” where phospholipid were provided from the krill oil used.

The microdiet was prepared according to Liu et al. (2002) as follows: the krill meal was mixed with the water-soluble ingredients (attractants, minerals and water-soluble vitamins). Oil and fat-soluble vitamins were mixed and blended with the dry ingredients. Finally, gelatine dissolved in warm water was added to the mix. The paste was pelleted and dried at 38° C for 24 h. The final pellets were ground and sieved in two different particle

sizes (120-250 and 250-500 μm). Diets were kept at 4° C during the feeding period. Proximate composition and fatty acids levels were analysed for each diet prior to the start of the trial (Table 1 and 2). Fatty acid profile was similar in all the experimental diets excepting for diet “KRILL”, which showed higher amounts of EPA (16.0 %) and DHA (8 %) than the other diets, what in turn increased total n-3 PUFA (Table 2). On the other hand, total n-6 PUFA was lower in KRILL, mainly due to the higher amount of linoleic acid in the diets with fish oil (7.2 % versus 4.1 %). Fish larvae were fed each 45 min daily from 8:00 to 20:00 with 3, 3.5 and 4 g tank⁻¹, during the first, second and third week respectively.

2.3. Sample collection

Samplings were performed at 30 and 44 dph. At the beginning of the experiment (30 dph) 100 larvae were sacrificed with an overdose of anaesthetic (clove oil; Guinama, Valencia, Spain) and fixed in 4 % buffered formalin for histological analysis. After two weeks (44 dph) 70 larvae per tank were sacrificed with clove oil and kept in ice during the sampling. 40 larvae were measured for total length (TL) using a profile projector (Mitutoyo PJ- 3000A, Kanagawa, Japan) and fixed in 4 % buffered formalin for histological analysis (120 larvae per diet). The remaining 30 larvae were collected to determine dry weight at each sampling point. At 44 dph all remaining larvae were collected for biochemical and TBARs analysis and stored at -80 °C until analysis.

2.4. Growth and survival

Larvae were sampled and measured for dry weight (100 °C for 24 h) and total length at the end of the experiment (44 dph). Final survival was determined at 44 dph by counting the remaining alive larvae in experimental tanks. Performance parameters were calculated according to the following equations: Survival (%) = 100*(final number fish - initial number fish)/initial number fish; SGR (specific growth rate) = 100*(ln final mean weight - ln initial mean weight)/number of days.

2.5. Histopathology

Formalin fixed samples were dehydrated in a series of different concentrations of ethanol and embedded in a paraffin block. The samples were cut at 4 µm on a microtome, fixed to the microscope slide and finally stained with haematoxylin and eosin (H&E), Ziehl-Neelsen (ZN) (Martoja and Martoja-Pearson, 1970), Fite-Faraco method (Fite et al., 1947) and Gram stain (Gregersen, 1978). Then, the samples were used for histopathological evaluation, analysing all tissues and focusing especially, in liver, kidney and heart, given that these organs are the main affected by granulomas (Ruiz et al., 2018a).

2.6. Biochemical analysis

Larvae and diet biochemical composition analysis were conducted following standard procedures. Lipids of larvae and feeds were extracted with a chloroform-methanol (2:1 v/v) mixture as described by Folch et al. (1957). Protein content (Kjeldahl method), dry matter and ash were determined in feeds according to AOAC (2010).

Fatty acids from total lipids were prepared by transmethylation as described by Christie (1982). Fatty acid methyl esters (FAMES) were separated and quantified by gas-liquid chromatography following the conditions described by Izquierdo et al. (1992). Lipid susceptibility to oxidation was estimated using the peroxidation index (PIn) with following formula: $PIn = 0.025 \times (\text{percentage of monoenoics}) + 1 \times (\text{percentage of dienoics}) + 2 \times (\text{percentage of trienoics}) + 4 \times (\text{percentage of tetraenoics}) + 6 \times (\text{percentage of pentaenoics}) + 8 \times (\text{percentage of hexaenoics})$ (Witting and Horwitt, 1964).

Thiobarbituric acid reactive substances (TBARs) were measured in triplicate from extracted total lipids (10 mg/ml) according to Burk et al. (1980). Firstly, 50 µl of 0.2 % (w/v) BHT in ethanol were added to 2 mg of lipid followed by 0.5 ml of 1 % (w/v) TBA and 0.5 ml of 10 % (w/v) trichloroacetic acid, all solutions freshly prepared. Samples were vortexed in stoppered test tubes and heated in darkness at 100 °C for 20 min. Then, samples were cooled in ice for 5 min and particulate matter was removed by centrifugation at 2,000 g (Sigma 4K15, Osterode am Harz, Germany) for 5 min. The supernatant was read in a spectrophotometer (Evolution 300, Thermo Scientific, Cheshire, UK) at 532 nm and recorded against a blank sample. The concentration of TBA-malondialdehyde (MDA) was

expressed as $\mu\text{mol MDA per g of tissue}$ and was calculated using the extinction coefficient $0.156 \mu\text{M}^{-1} \text{ cm}^{-1}$.

The concentration of vitamin E was determined in diets. Samples were weighed, homogenized in ethanolic pyrogallol and saponified as described McMurray et al., 1980. HPLC analysis was performed using 150 x 4.60 mm, 5 μm reverse-phase Luna and C18 column (Phenomenox, CA, USA). The mobile phase was methanol:ultrapure water (98:2 v/v) with a flow rate of 1.0 ml min^{-1} at ambient temperature. Samples were injected (50 μl) in a high performance liquid chromatograph (HPLC) with UV detection at a wavelength of 293 nm to determine the vitamin E using (+)- α -tocopherol (Sigma-Aldrich) as the external standard.

The concentration of vitamin C was determined in the experimental feeds as described by Betancor et al. (2012). Samples were weighed, homogenised and dissolved in 0.4 M phosphate buffer (adjusted to pH 3.0 with phosphoric acid). The samples were centrifuged at 3.000 rpm, supernatants removed and filtered through a disposable 0.45 μm filter and stored at 4° C until the measurement in a HPLC with UV detection. The determination of vitamin C concentration was achieved by comparison with tris (cyclohexylammonium) ascorbic acid-2-phosphate (Sigma-Aldrich) as the external standard.

2.7. Statistical analysis

All statistical analyses were done with Statgraphics (Statgraphics Centurion XVI version 16.1.03 for Windows; Graphic Software Systems, Inc. USA). Survival, growth, percentage of larvae with granulomas and biochemical analysis were tested for normality with the Kolmogorov Smirnov test and homogeneity of variance was performed with the Levene test. With the variables that satisfied the normality and homogeneity was carried out a parametric one-way (ANOVA) and Tukey test post-hoc test. Correlations were analysed with Pearson's correlation coefficient. A significance level of 0.05 was used.

3. Results

3.1. Growth and survival

All experimental diets were well accepted by larvae. Final total length, dry weight and survival were not significant different among larvae fed the different experimental feeds at the end of the feeding trial (44 dph). The average final total length was 25.8 ± 0.4 mm, dry weight 17.5 ± 1.3 mg, survival 20.1 ± 0.5 % and SGR 17.1 ± 1.2 % (Table 3).

3.2. Histopathology

At the beginning of the experiment (30 dph) no granulomas were observed at the microscopic evaluation. Nevertheless, after 14 days (44 dph) significant differences were found in the percentage of larvae with granulomas among diets, being higher in larvae fed diets C-, Krill and EC (40/100, 400/1,000 and 200/500 mg kg⁻¹ of vitamin E and C, respectively) followed by diet C+ (400/1,000 mg kg⁻¹ of vitamin E and C, respectively) (Figure 1). No granulomas were observed in any larvae fed with the highest levels of vitamin E and C (800/2,000 mg kg⁻¹). Kidney was the main affected tissue with granulomas (86.7 % of fish with granulomas), followed by liver (13.3 % of fish with granulomas) (Figure 2).

There was a strong and significant negative correlation between the percentage of larvae with granulomas and dietary concentration of vitamin E ($y = -0.0098x + 11.174$, $R^2 = 0.8766$; Pearson's correlation coefficient (r) = -0.93) and vitamin C ($y = -0.0022x + 6.4777$, $R^2 = 0.9278$; Pearson's correlation coefficient (r) = -0.96) (Figure 3). The TBARs content was highly correlated with the appearance of granulomas ($R^2=0.892$, $y=0.0446x+0.0756$).

All the specific stainings (Ziehl-Neelsen, Fite-Faraco and Gram stain) were negative, discarding a possible infectious origin of the granulomas (Supplementary Figure 1).

The histopathological evaluation revealed granulomas in different stages of development (Supplementary Figure 2) as described by Ruiz et al. (2018a) in on-growing meagre. At initial stages, granulomas were observed as isolated and irregular aggregated of macrophages (Supplementary Figure 2a) that later were forming concentric layers

(Supplementary Figure 2b). These aggregated progressively lead to a necrotic centre with external layers of fibrocytes (Supplementary Figure 2c). However, final stages of development, in which the granuloma is completely composed of laminar material, were not observed.

3.3. Biochemical analysis

3.3.1. Whole larvae proximal composition and fatty acid profile

Dietary treatment did not affect larvae whole body proximate composition after 14 days of feeding, with a protein content averaging 11 % and lipid exceeding 2 % among larvae fed the different dietary treatments (Table 4). The substitution of krill oil by fish oil significantly increased the levels of eicosapentaenoic acid (EPA, 16.6 vs 13.7 %) and docosahexaenoic acid (DHA, 17.63 vs 16.05 %) in meagre larvae at the end of the feeding trial, compared with the larvae fed with the other diets (EPA ~ 13.7 % and DHA ~ 16.1 %) (Table 5). Furthermore, the addition of krill oil significantly increased the peroxidation index in the larvae (275.3 vs 245.7) (Table 5) and the concentration of saturated fatty acids (31.2 vs 29.0 %), n-3 PUFA (38.2 vs 34.3 %) and n-3 LC-PUFA (35.6 vs 31.6 %) (Table 5). Larvae fed fish oil diets showed significant higher concentration of oleic acid, linoleic acid, monosaturated fatty acids, n-6 and n-9 PUFA regardless dietary levels of vitamin E and C (Table 5).

3.3.2. TBARs content

The level of lipid peroxides, as indicated by TBARs content ($\mu\text{mol g}^{-1}$ larval tissues), was significantly lower in those larvae fed diets with the highest levels of vitamin E and C (Table 4).

4. Discussion

It has previously been shown that the co-feeding with rotifer and *Artemia* enriched with Easy DHA Selco prior to eating an inert commercial microdiet prevented the appearance of granulomas in meagre larvae (Ruiz et al., 2018b). Consistently, no granulomas were observed at 30 dph in the present trial after following the same feeding sequencing and enrichment protocol what seems to reinforce the role of nutrition as the main trigger in the appearance of systemic granulomatosis. The results of the present trial

showed that the dietary addition of different levels of vitamin E (40, 200, 400 and 800 mg kg⁻¹) and C (100, 500, 1,000 and 2,000 mg kg⁻¹) did not affect meagre larvae performance in terms of growth, length, survival and SGR at 44 dph. However, granulomas were observed in larvae fed with low levels of vitamin E and C (from 40/100 to 400/1,000 mg kg⁻¹, vitamin E/C). The results suggest that low levels of vitamin E and C (40 and 100 mg kg⁻¹, respectively) probably fulfilled the requirement for normal growth what explains the lack of differences in terms of fish performance among larvae fed the different dietary treatments but were not enough to prevent systemic granulomatosis. On this matter, a strong negative correlation was observed between the dietary levels of vitamin E ($y = -0.0098x + 11.174$, $R^2 = 0.8766$) and vitamin C ($y = -0.0022x + 6.4777$, $R^2 = 0.9278$) and the incidence of granulomas. Little is known about requirements of vitamin E and C in meagre larvae. Only El Kertaoui et al. (2017) observed that high levels (1,500 and 1,800 mg kg⁻¹ of vitamin E and C, respectively) were required to improve growth and antioxidant defenses in meagre larvae at 28 dph. It is well known that the requirement for antioxidant vitamins is conditioned by the dietary fatty acids content. In this regard, all the experimental microdiets contained a sufficient amount of essential fatty acids for most marine fish species, which require at least 2 % EPA and DHA (NRC, 2011). Nevertheless, those larvae fed with higher amounts of DHA and EPA together with low dietary vitamin E and C (diet Krill) presented high incidence of granulomas, suggesting an imbalance between prooxidant and antioxidant nutrients. Accordingly, TBARs content, an indicator of lipid oxidation, was affected by the dietary inclusion of vitamin E and C, with the high supplementation of vitamin E (800 mg kg⁻¹) and C (2,000 mg kg⁻¹) significantly reducing TBARs values. Indeed, TBARs contents were highly correlated with the appearance of granulomas ($R^2=0.892$, $y=0.0446x+0.0756$). Therefore, adequate dietary levels of vitamins E and C seem to mitigate the appearance of systemic granulomatosis in meagre larvae, probably due to the decrease of the oxidation rate.

Vitamin E together with vitamin C are strong antioxidants in tissues, being able to neutralize reactive oxygen species (ROS) (Montero et al., 1999; Ai et al., 2006; Betancor et al., 2012; Gao et al., 2014) and increase the protection against lipid peroxidation (Lee and Dabrowski, 2003). The oxidative stress has been related with some diseases (Kawatsu, 1969; Cowey et al., 1984; Sakai et al., 1989; Watanabe et al., 1989; Sies et al., 1992;

Padayatty and Levine, 2001; Lewis-McCrea and Lall, 2007), therefore it is feasible to think that granulomas could also be originated by an oxidative imbalance. Lipid peroxidation contributes to the inflammatory response (Morita et al., 2016). Granuloma formation is an inflammatory response, and is composed basically by macrophages, lymphocytes and fibrocytes, being its appearance not necessarily associated with infectious diseases. This inflammation can occur in blood vessels (Petersen and Smith, 2013; Hilhorst et al., 2014). In this sense, in the present and previous studies (Ruiz et al., 2018a) irregular aggregates of cells and granulomas have been observed surrounding blood vessels, which suggests that granulomas could have a vascular origin. Vitamin C has been related with the synthesis of collagen, an important protein involved in the generation of blood vessels (Lim and Lovell, 1978; Nusgen et al., 2001). Besides, vitamins C and E are involved in the prevention of endothelial dysfunction and the prevention of oxidative stress (Riitta et al., 2003; Engler et al., 2003). In this sense, an imbalance between ROS and antioxidants could be happening in larvae fed with low addition of vitamin E and C, as indicated by TBARs values, which could lead to inflammatory response in blood vessel with the subsequent macrophages infiltration and formation of granulomas. Limited information is available on the effect of antioxidant vitamins in the formation of granulomas. In other fish species vitamin C deficiency has been related to precipitation of tyrosine in tissues, being the origin of granulomas, in species such as sea bream and turbot (Baudin-Laurencin et al., 1989; Coustans et al., 1990; Alexis et al., 1997). In agreement, a previous study showed that the dietary increase of vitamins E and C lead to a reduction in the percentage of granulomas in liver and heart of juvenile meagre together with a decrease in TBARs contents (Ruiz et al., 2018ab), what indicates less lipid peroxidation.

The substitution of fish oil by krill oil significantly increased the levels of eicosapentaenoic acid (EPA, 16.6 %) and docosahexaenoic acid (DHA, 17.6 %) in meagre larvae with 44 dph, compared with the larvae fed the other diets (EPA ~ 13.7 and DHA ~ 16.1 %). This difference in the levels of n-3 LC-PUFA seemed to have an impact on the TBARs content which in turn translated into a higher incidence of granulomas compared to larvae fed fish oil in combination with the same dietary levels of antioxidant vitamins (Diet C+, 400 and 1,000 mg kg⁻¹ vitamin E and C, respectively). Apart from being an excellent source of EPA and DHA, krill oil is rich in phospholipids and particularly

phosphatidylcholine (Winther et al., 2011). Phospholipids have been described to have a stronger biological effect than triglycerides, because they can be more rapidly digested and are more effectively incorporated to the tissues than triglycerides (Ackman and Ratnayake 1989), can act as ligands for nuclear receptor (Li et al., 2005; Chakravarthy et al., 2009), are involved in the steroidogenesis and cholesterol metabolism, and have been shown to augment the bioavailability of DHA and EPA (Amate et al., 2001; Cansell et al., 2003; Cansell et al., 2009). Despite of the high phospholipid level provided by the krill oil, it could not prevent the appearance of granulomas, needing supplementation with higher levels of vitamin E and C (over 400 and 1,000 mg kg⁻¹, respectively) in order to inhibit its appearance. However, the percentage of granulomas was significantly higher in larvae fed diet “krill” than those larvae fed diet “C+”, although both diets contained the same levels of vitamin E (844 and 859 mg kg⁻¹, respectively) and C (1,460 and 1,450 mg kg⁻¹, respectively) were roughly the same. This could be related to the higher EPA and DHA contents (therefore, higher peroxidation index) found in larvae fed diet “krill”, what suggests that the balance between prooxidant and antioxidant nutrients is disturbed in favour of prooxidants. In this point, it should be noted that the higher peroxidation index should be correlated to higher TBARs values. Nevertheless, larvae fed diet “krill” were not different to those of fish fed fish oil (C+). This could be due the fact that EPA and DHA are in phospholipid forms and were more protected in the krill diet, while in the diet with fish oil they were in triglycerides, being more susceptible to oxidation. Moreover, although krill oil contains antioxidants, mainly astaxanthin (Tou et al., 2007), these were no able to prevent the appearance of granulomas. These results suggest that the appearance of granulomas is more related to the supplementation of different levels of vitamin E and C more than to the source of dietary fatty acids. In fact, in a previous study the appearance of granulomas in juvenile meagre was modulated by the inclusion of different levels of the antioxidants vitamins E and C (Ruiz et al., 2018a).

Concluding, the supplementation of vitamin E and C at 40 and 100 mg kg⁻¹ respectively is adequate to ensure good meagre larvae performance. However, these vitamin levels might not be enough to prevent the appearance of systemic granulomatosis, as indicated by the strong negative correlation between dietary vitamin E and C contents and the prevalence of granulomas and TBARs values. Levels of dietary vitamin E and C of

1,082 and 2,910 mg kg⁻¹ (Diet EECC) completely prevented the appearance of granulomas. The substitution of fish oil by krill oil was enough to the correct growth of meagre larvae but increased the percentage of granulomas and the peroxidation index. Therefore, it has been demonstrated in the present and previous studies (Ruiz et al., 2018b) that systemic granulomatosis can be completely mitigated in meagre larvae by controlling feeding sequence as well as levels of antioxidant nutrients.

5. Acknowledgements

This study was funded by the project “Exploring the biological and socio-economic potential of new/emerging candidate fish species for expansion of the European aquaculture industry (DIVERSIFY)” of the European Commission; Directorate-general for research and Innovation, project no. FP7-KBBE-2013-7, GRANT AGREEMENT NUMBER 603121.

6. References

- Ackman, R.G., Ratnayake, W.M.N., 1989. Fish oils, seal oils, esters and acids -are all forms of T-3 intake equal? In: Health Effect of Fish and Fish Oils (Chandra, R.K. Eds.), ARTS Biomedical, Newfoundland, Canada, pp. 373-393.
- Ai, Q., Mai, K., Tan, B., Xu, W., Zhang, W., Ma, H., Liufu, Z., 2006. Effects of dietary vitamin C on survival, growth, and immunity of large yellow croaker, *Pseudosciaena crocea*. Aquaculture. 261, 327–336. <https://doi.org/10.1016/j.aquaculture.2006.07.027>
- Alexis, M.N., Karanikolas, K.K., Richards, R.H., 1997. Pathological findings owing to the lack of ascorbic acid in cultured gilthead bream (*Sparus aurata* L.). Aquaculture. 151, 209-218. [https://doi.org/10.1016/S0044-8486\(96\)01475-5](https://doi.org/10.1016/S0044-8486(96)01475-5)
- Amate, L., Gil, A., Ramirez, M., 2001. Feeding infant piglets formula with long-chain polyunsaturated fatty acids as triacylglycerols or phospholipids influences the distribution of these fatty acids in plasma lipoprotein fractions. J. Nutr. 131, 1250-1255. <https://doi.org/10.1093/jn/131.4.1250>
- AOAC, 2010. Official Methods of Analysis, 17th Ed. Association of Official Analytical Chemists, Washington, D. C., U.S.A.
- Baudin-Laurencin, F., Messenger, J.L., Stephan, G., 1989. Two examples of nutritional pathology related to vitamin E and C deficiencies, in: Advances in tropical aquaculture, Tahiti, 171-181.
- Betancor, M.B., Caballero, M.J., Terova, G., Corà, S., Saleh, R., Benítez-Santana, T., Bell, J.G., Hernández-Cruz, C.M., Izquierdo, M., 2012. Vitamin C Enhances Vitamin E Status and Reduces Oxidative Stress Indicators in Sea Bass Larvae Fed High DHA Microdiets. Lipids. 47, 1193–1207. <https://doi.org/10.1007/s11745-012-3730-x>
- Burk, R.F., Trumble, M.J., Lawrence, R.A., 1980. Rat hepatic cytosolic GSH-dependent enzyme protection against lipid peroxidation in the NADPH microsomal lipid peroxidation system. Biochim. Biophys. Acta. 618, 35-41. [https://doi.org/10.1016/0005-2760\(80\)90051-X](https://doi.org/10.1016/0005-2760(80)90051-X)

451 Cansell, M., Nacka, F., Combe, N., 2003. Marine lipid-based liposomes increase in vivo FA bioavailability. *Lipids*. 38,
452 551-559. <https://doi.org/10.1007/s11745-003-1341-0>

453 Cansell, M.S., Battin, A., Degrace, P., Gresti, J., Clouet, P., 2009. Early dissimilar fates of liver eicosapentaenoic Acid in
454 rats fed liposomes or fish oil and gene expression related to lipid metabolism. *Lipids*. 44, 237-247.
455 <https://doi.org/10.1007/s11745-008-3279-x>

456 Carvalho, M., Castro, P., Peres, H., Fontanillas, R., Rosenlund, G., Monter, D., Acosta, F., Robaina, L., Izquierdo, M.,
457 2018. Effect of increasing dietary levels of n-3 long-chain polyunsaturated fatty acids on liver composition and
458 histopathology of meagre (*Argyrosomus regius*, Asso 1801) fingerlings. 18th International Symposium on Fish Nutrition
459 and Feeding, 3-7 June 2018, Las Palmas de Gran Canaria, Spain.

460 Chakravarthy, M.V., Lodhi, I.J., Yin, L., Malapaka, R.R.V., Xu, H.E., 2009. Identification of a physiologically relevant
461 endogenous ligand for PPAR α in liver. *Cell*. 138, 476-488. <https://doi.org/10.1016/j.cell.2009.05.036>

462 Christie, W.W., 1982 . *Lipid Analysis*, 2nd Ed. Pergamon, Oxford.

463 Cotou, E., Fountoulaki, E., Hager-Theodorides, A.L., Theodorou, G., Tsertou, M., Loukanari T., Katharios, P., Kounna,
464 C.H., 2016. The effects of dietary inclusions of vitamin D3 associated to CYP27A1, antioxidant enzymes and non-
465 infectious systemic granulomatosis in meagre (*Argyrosomus regius*). International Symposium on Fish Nutrition and
466 Feeding, 5-10, 2016, Sun Valley, Idaho.

467 Coustans, M.F., Guillaume, J., Metailler, R., Dugornay, O., Messenger, J.L., 1990. An ascorbic acid deficiency on
468 tyrosinemia and renal granulomatous disease in turbot (*Scophthalmus maximus*) interaction with a slight
469 polyhypovitaminosis. *Comparative Biochemistry and Physiology Part A: Physiology*. 97, 145-152.
470 [https://doi.org/10.1016/0300-9629\(90\)90161-K](https://doi.org/10.1016/0300-9629(90)90161-K)

471 Cowey, C.B., Degener, E., Tacon, A.G.J., Youngson, A., Bell, J.G., 1984. The effects of vitamin E and oxidised fish oil
472 on the nutrition of rainbow trout (*Salmo gairdneri*) grown at natural, varying water temperature. *Br. J. Nutr.* 51, 443-451.
473 <https://doi.org/10.1079/BJN19840050>

474 Dabrowski, K., 1992. Ascorbate concentration in fish ontogeny. *J. Fish Biol.* 40, 273–279. <https://doi.org/10.1111/j.1095-8649.1992.tb02572.x>

476 El Kertaoui, N., Hernández- Cruz, C.M., Montero, D., Caballero, M.J., Saleh, R., Afonso, J.M., Izquierdo, M., 2017. The
477 importance of dietary HUFA for meagre larvae (*Argyrosomus regius*; Asso, 1801) and its relation with antioxidant
478 vitamins E and C. *Aquac. Res.* 48, 419-433. <https://doi.org/10.1111/are.12890>

479 Fite, G.L., Cambre, F.J., Turner, M.H., 1947. Procedures for demonstrating lepra bacilli in paraffin sections. *Arch. Pathol.*
480 43, 624-625.

481 Folch, J.M., Lees M., Stanley Sloane, G.H., 1957. A simple method for the isolation and purification of total lipids from
482 the animal tissues. *J. Biol. Chem.* 226, 497-509.

483 Gao J., Koshio, S., Ishikawa, M., Yokoyama, S., Edward, R., 2014. Interactive effects of vitamin C and E
484 supplementation on growth performance, fatty acid composition and reduction of oxidative stress in juvenile Japanese
485 flounder *Paralichthys olivaceus* fed dietary oxidized fish oil. *Aquaculture*. 422-423, 84-90.
486 <https://doi.org/10.1016/j.aquaculture.2013.11.031>

487 Ghittino, C., Manuali, E., Latini, M., Agnetti, F., Rogato, F., Agonini, R., Colussi, S., Prearo, M., 2004. Caso di
488 granulomatosi sistemica in ombrina boccardoro (*Argyrosomus regius*) e raffronto con le lesioni istologiche presenti
489 nell'orata. *Ittiopatologia*. 1, 59-67.
490

491 Goldsmith, L.A., 1978. Molecular biology and molecular pathology of a newly described molecular disease. Tyrosinemia
492 II (the Richner - Hanhart syndrome). *Experimental cell biology*. 46, 96-113.

493 Gregersen, T., 1978. Rapid method for distinction of gram-negative from gram-positive bacteria. Eur. J. Appl. Microbiol.
494 Biotechnol. 5, 123-127. <https://doi.org/10.1007/BF00498806>

495 Hamre, K., 2011. Metabolism, interactions, requirements and functions of vitamin E in fish. Aquacult. Nutr. 17, 98-115.

496 Hilhorst, M., Shirai, T., Berry, G., Goronzy, J.J., Weyand, C.M., 2014. T Cell–Macrophage Interactions and Granuloma
497 Formation in Vasculitis. Front. Immunol. 5, 432. <https://doi.org/10.3389/fimmu.2014.00432>

498 Izquierdo, M.S., 1996. Essential fatty acid requirements of cultured marine fish species. Aquacult. Nutr. 2, 193-191.
499 <https://doi.org/10.1111/j.1365-2095.1996.tb00058.x>

500

501 Izquierdo, M.S., Koven, W., 2011. Lipids, in: Holt J. (Eds.), Larval Fish Nutrition., Wiley-Blackwell, Chichester, UK, pp.
502 47-84.

503

504 Izquierdo, M.S., Arakawa, T., Takeuchi, T., Haroun, R., Watanabe, T., 1992. Effect of n-3 HUFA levels in Artemia on
505 growth of larval Japanese flounder (*Paralichthys olivaceous*). Aquaculture. 105, 73-82. [https://doi.org/10.1016/0044-](https://doi.org/10.1016/0044-8486(92)90163-F)
506 8486(92)90163-F

507

508 Kawatsu, H., 1969. Studies on the anemia of fish-III. An example of macrocyticanemia found in brook trout, *Salvelinus*
509 *fontinalis*. Bulletin of Freshwater Fisheries Research Laboratory. 19, 161-167.

510

511 Lee, K., Dabrowski, K., 2003. Interaction between vitamins C and E affects their tissue concentrations, growth, lipid
512 oxidation, and deficiency symptoms in yellow perch (*Perca flavescens*). Br. J. Nutr. 89, 589-596.
513 <https://doi.org/10.1079/BJN2003819>

514 Lewis-McCrea, L.M., Lall, S.P., 2007. Effects of moderately oxidized dietary lipid and the role of vitamin E on the
515 development of skeletal abnormalities in juvenile atlantic halibut (*Hippoglossus hippoglossus*). Aquaculture. 262, 142-
516 155. <https://doi.org/10.1016/j.aquaculture.2006.09.024>

517

518 Li, Y., Choi, M., Cavey, G., Daugherty, J., Suino, K., 2005. Crystallographic identification and functional characterization
519 of phospholipids as ligands for the orphan nuclear receptor steroidogenic factor-1. Mol. Cell. 17, 491-502.
520 <https://doi.org/10.1016/j.molcel.2005.02.002>

521

522 Lim, C., Lovell, R.T., 1978. Pathology of the vitamin C deficiency syndrome in channel catfish (*Ictalurus punctatus*). J.
523 Nutr. 108, 1137-1146. <https://doi.org/10.1093/jn/108.7.1137>

524

525 Liu, J., Caballero, M.J., Izquierdo, M.S., El-Sayed Ali, T., Hernández-Cruz, C.M., Valencia, A., Fernández-Palacios, H.,
526 2002. Necessity of dietary lecithin and eicosapentaenoic acid for growth, survival, stress resistance and lipoprotein
527 formation in gilthead sea bream *Sparus aurata*. Fisheries Sci. 6, 1165-1172. [https://doi.org/10.1046/j.1444-](https://doi.org/10.1046/j.1444-2906.2002.00551.x)
528 2906.2002.00551.x

529

530 Engler, M.M., Engler, M.B., Mallor, M.J., Chiu, E.Y., Schloetter, M.C., Paul, S.M., Stuehlinger, M., Lin, K.Y., Cooke,
531 J.P., Morrow, J.D., Ridker, P.M., Rifai, N., Miller, E., Witztum, J.L., Mietus-Snyder, M., 2003. Antioxidant vitamins C
532 and E improve endothelial function in children with hyperlipidemia. Circulation. 108, 1059-1063.
533 <https://doi.org/10.1161/01.CIR.0000086345.09861.A0>

534 Martoja, R., Martoja-Pearson, M., 1970. Técnicas de Histología Animal, 1º Ed. Toray-Masson S.A., Barcelona.

535

536 McMurray, C.H., Blanchflower, W.J., Rice, D.A., 1980. Influence of extraction techniques on determination of α -
537 tocopherol in animal feedstuffs. J. AOAC Int. 63, 1258–1261.

538

539 Montero, D., Marrero, M., Izquierdo, M.S., Robaina, L., Vergara, J.M., Tort, L., 1999. Effect of vitamins E and C dietary
540 supplementation on some immune parameters of gilthead seabream (*Sparus aurata*) juveniles subjected to crowding
541 stress. Aquaculture. 171, 269-78. [https://doi.org/10.1016/S0044-8486\(98\)00387-1](https://doi.org/10.1016/S0044-8486(98)00387-1)

542 Morita, M., Naito, Y., Yoshikawa, T., Niki, E., 2016. Plasma lipid oxidation induced by peroxynitrite, hypochlorite,
543 lipoxygenase and peroxy radicals and its inhibition by antioxidants as assessed by diphenyl-1-pyrenylphosphine. *Redox*
544 *Biol.* 8, 127–135. <https://doi.org/10.1016/j.redox.2016.01.005>

545 NRC (2011) Nutrient Requirements of Fish and Shrimp. National Academic Press, Washington D.C.
546 <https://doi.org/10.17226/13039>

547 Nusgens, B.V., Colige, A.C., Lambert, C.A., Lapière, C.M., Humbert, P., Rougier, A., Haftek, M., Richard, A., Creidi, P.,
548 2001. Topically Applied Vitamin C Enhances the mRNA Level of Collagens I and III, Their Processing Enzymes and
549 Tissue Inhibitor of Matrix Metalloproteinase 1 in the Human Dermis. *J. Invest. Dermatol.* 116, 853-859.
550 <https://doi.org/10.1046/j.0022-202x.2001.01362.x>

551 Padayatty, S.J., Levine, M., 2001. New insights into the physiology and pharmacology of vitamin C. *CMAJ* 164, 353–
552 355.

553 Paperna, I., Harrison, J.G., Kissil, G.W., 1980. Pathology and histopathology of a systemic granuloma in *Sparus aurata*
554 (L.) cultured in the Gulf of Aqaba. *J. Fish Dis.* 3, 213-221.

555 Petersen, H., Smith, A., 2013. The role of the innate immune system in granulomatous disorders. *Front. Immunol.*
556 <https://doi.org/10.3389/fimmu.2013.00120>

557 Riitta, M.S., Kristiina, N., Jari, K., Elina, P., Sari, V., Tiina, H.R., Tomi-Pekka, T., Veli-Pekka, V., Ulla, R., Hanna-Maari,
558 L., Meri, V., Jukka, T.S., Henrik, E.P., 2003. Six-Year Effect of Combined Vitamin C and E Supplementation on
559 Atherosclerotic Progression. *Circulation.* 107, 947-953. <https://doi.org/10.1161/01.CIR.0000050626.25057.51>

560 Rodríguez, C., Pérez, J.A., Lorenzo, A., Izquierdo, M.S., Cejas, J.R., 1994. n-3 HUFA requirement of larval gilthead
561 seabream *Sparus aurata* when using high levels of eicosapentaenoic acid. *Comparative Biochemistry and Physiology Part*
562 *A: Physiology.* 4, 693-698. [https://doi.org/10.1016/0300-9629\(94\)90371-9](https://doi.org/10.1016/0300-9629(94)90371-9)

563 Ruiz, M.A., Betancor, M.B., Robaina, L., Montero, D., Hernández-Cruz, C.M., Izquierdo, M., Rosenlund, G., Fontanillas,
564 R., Caballero, M.J., 2018a. Dietary combination of vitamin E, C and K affects growth, antioxidant activity, and the
565 incidence of systemic granulomatosis in meagre (*Argyrosomus regius*). *Aquaculture.* 498, 606-620.
566 <https://doi.org/10.1016/j.aquaculture.2018.08.078>

567
568 Ruiz, M.A., Hernández-Cruz, C.M., Caballero M.J., Fernández-Palacios, H., Saleh, R., Izquierdo, M.S., Betancor, M.B.,
569 2018b. Incidence of systemic granulomatosis is modulated by the feeding sequence and type of enrichment in meagre
570 (*Argyrosomus regius*). *Aquacult. Res.* 50, 284-295. <https://doi.org/10.1111/are.13896>

571 Sakai, T., Murata, H., Endo, M., Yamauchi, K., Tabata, N., Fukudome, M., 1989. 2-Thiobarbituric acid values and
572 contents of α -tocopherol and bile pigments in the liver and muscle of jaundiced yellowtail, *Seriola quinqueradiata*. *Agric.*
573 *Biol. Chem.* 53, 1739-1740. <https://doi.org/10.1080/00021369.1989.10869531>

574 Salhi, M., Izquierdo, M.S., Hernández-Cruz, C.M., Socorro, J., Fernández-Palacios, H., 1997. The improved incorporation
575 of polyunsaturated fatty acids and changes in liver structure in larval gilthead seabream fed on microdiets. *J. Fish Biol.* 51,
576 869-879. <https://doi.org/10.1111/j.1095-8649.1997.tb01526.x>

577
578
579 Sies, H., Stahl, W., Sundquist, A.R., 1992. Antioxidant functions of vitamins. Vitamins E and C, beta-carotene and other
580 carotenoids. *Ann. NY Acad. Sci.* 30, 7-20. <https://doi.org/10.1111/j.1749-6632.1992.tb17085.x>

581 Tou, J.C., Jaczynski, J., Chen, Y.C., 2007. Krill for human consumption: nutritional value and potential health benefits.
582 *Nutr. Rev.* 65, 63-77. <https://doi.org/10.1111/j.1753-4887.2007.tb00283.x>

583 Ulven, S.M., Holven, K.B., 2015. Comparison of bioavailability of krill oil versus fish oil and health effect. *Vasc. Health*
584 *Risk. Manag.* 11, 511-524. doi:10.2147/VHRM.S85165

Watanabe, T., Izquierdo, M.S., Takeuchi, T., Satoh, S., Kitajima, C., 1989. Comparison between eicosapentaenoic and docosahexaenoic acids in terms of essential fatty acids efficacy in larval red seabream. *Nippon Suissan Gakk.* 5, 1635-1640. <https://doi.org/10.2331/suisan.55.1635>

Winther, B., Hoem, N., Berge, K., Reutsaet, L., 2011. Elucidation of phosphatidylcholine composition in krill oil extracted from *Euphausia superba*. *Lipids.* 46, 25–36. <https://doi.org/10.1007/s11745-010-3472-6>

Witting, L.A., Horwitt, M.K., 1964. Effect of degree of fatty acid unsaturation in tocopherol deficiency-induced creatinuria. *J. Nutr.* 82, 19–33. <https://doi.org/10.1093/jn/82.1.19>

Figure legends

Figure 1. Incidence of granulomas (%) in meagre larvae at the end of the dietary trial (44 dph). Each value represents mean \pm SD (n= 120).

Figure 2. Percentage of affected organs with granulomas in meagre larvae of 44 dph fed different levels of vitamin E and C.

Figure 3. Effect of dietary vitamin E and C on percentage of affected meagre larvae with granulomas at 44 dph.

Supplementary Figure 1. Negative results in granulomas for specific stains. A) Ziehl-Neelsen, B) Gram stain and C) Fite-Faraco stain in kidney.

Supplementary Figure 2. Granulomas at different stages of development in kidney of meagre larvae (44 dph) at the end of the experimental trial. **A)** Irregular aggregated of macrophages. **B)** Concentric layers of macrophages and some lymphocytes. **C)** Necrotic center surrounded by layers of macrophages and an outer layer of fibrocytes.

Tables

Table 1. Formulation and analysed proximate composition of diets fed to meagre larvae from 30 to 44 dph, containing different levels of vitamin E and C and either fish or krill oil as the lipid source.

Ingredient (%)	Diets				
	C+	C-	EC	EECC	Krill
Krill meal	74.47	74.60	74.54	74.33	75.47
Krill oil	-	-	-	-	6.00
Gelatin ¹	3.00	3.00	3.00	3.00	3.00
Fish oil	7.00	7.00	7.00	7.00	-
Soy lecithin ²	2.00	2.00	2.00	2.00	2.00
Vitamin E ³	0.04	0.004	0.02	0.08	0.04
Vitamin C ³	0.10	0.01	0.05	0.20	0.10
Mineral Premix ⁴	4.70	4.70	4.70	4.70	4.70
Vitamin Premix ⁵	5.69	5.69	5.69	5.69	5.69
Attractant ⁶	3.00	3.00	3.00	3.00	3.00
Proximate composition					
Vitamin E (mg kg ⁻¹)	844.3	497.1	632.7	1082.3	859.8
Vitamin C (mg kg ⁻¹)	1460.8	153.1	758.5	2910.5	1450.2
Protein (%)	48.5	48.9	48.9	48.0	49.6
Lipid (%)	30.1	30.6	29.9	30.8	29.7
Moisture (%)	3.7	3.7	3.6	4.0	4.1
Ash (%)	11.8	11.7	11.9	11.9	11.9

¹Panreac, Barcelona, Spain. ²Acrofarma, Barcelona, Spain. ³ g · 100⁻¹, Vitamin E: α-tocopheryl acetate (Sigma-Aldrich, Madrid, Spain), Ascorbyl monophosphate ROVIMIX Stay-C-35 (Roche, Paris, France). ⁴Mineral premix supplied g per 100 g diet: NaCl 215.133 mg, MgSO₄ 7H₂O 677.545 mg, NaH₂PO₄ H₂O 381.453 mg, Ca(H₂PO₄) 2H₂O 671.610 mg, FeC₆H₅O₇ 146.884 mg, C₃H₅O₃ 1/2Ca 1,617.210 mg, Al₂(SO₄)₃ 6H₂O 0.693 mg, ZnSO₄ 7H₂O 14.837 mg, CuSO₄ 5H₂O 1.247 mg, MnSO₄ H₂O 2.998 mg, CoSO₄ 7H₂O 10.706 mg. ⁵Vitamin premix supplied per 100 g diet: cyanocobalamine 0.03 mg, astaxanthin 5.0 mg, folic acid 5.4 mg, pyridoxine-HCl 17.3 mg, thiamine 21.7 mg, riboflavin 72.5 mg, calcium-pantothenate 101.5 mg, p-aminobenzoic acid 145.0 mg, nicotinic acid 290.1 mg, myo-inositol 1450.9 mg, menadione 17.3 mg. ⁶Attractant premix supplied per 100 g diet: inosine-5-monophosphate 500.0 mg, betaine 660.0 mg, L-serine 170.0 mg, L-phenylalanine 250.0 mg, DL-alanine 500.0 mg, L-sodium aspartate 330.0 mg, L-valine 250.0 mg, glycine 170.0 mg. Proximate composition (%)

Table 2. Diets fatty acid composition (percentage of fatty acids) used for feeding meagre larvae fed from 30 to 44 days post hatching (dph) in the present trial.

<i>Fatty acids (%)</i>	Diets				
	C+	C-	EC	EECC	KRILL
14:0	7.1	7.1	7.2	7.2	9.7
16:0	19.4	19.4	19.6	19.7	22.6
18:0	2.1	2.1	2.1	2.1	1.7
20:0	0.1	0.1	0.1	0.1	0.1
Σ Saturated¹	29.3	29.3	29.6	29.7	34.8
16:1n-7	5.4	5.3	5.4	5.4	6.5
18:1n-9	20.4	20.5	20.5	20.3	13.3
18:1n-7	5.5	5.5	5.6	5.6	6.6
20:1n-7	1.8	1.8	1.8	1.8	1.0
22:1n-11	0.7	0.8	0.7	0.7	0.0
Σ Monosaturated²	35.6	35.8	35.8	35.5	29.5
18:2n-6	7.2	7.2	7.2	7.2	4.1
18:3n-6	0.1	0.1	0.1	0.1	0.1
20:2n-6	0.3	0.3	0.3	0.3	0.0
20:3n-6	0.1	0.1	0.1	0.1	0.0
20:4n-6	0.3	0.3	0.3	0.3	0.3
Σ n-6PUFA³	8.1	8.1	8.1	8.1	4.6
18:3n-3	1.9	1.9	1.9	1.9	1.0
18:4n-3	2.4	2.4	2.3	2.4	3.1
20:3n-3	0.1	0.1	0.1	0.1	0.1
20:4n-3	0.4	0.4	0.4	0.4	0.3
20:5n-3	12.1	12.0	11.9	12.1	16.0
22:5n-3	0.6	0.6	0.6	0.6	0.7
22:6n-3	7.4	7.4	7.3	7.2	8.0
Σ n-3PUFA⁴	25.0	24.8	24.6	24.7	28.8
(n-3+n-6) PUFA	33.2	32.9	32.7	32.8	33.4
Total n-3 LC-PUFA⁵	20.5	20.4	20.2	20.3	24.7
PIn	166.4	165.1	163.6	163.9	190.3

Data expressed as means of three technical replicates per batch of diet.¹Includes 15:0 and 17:0.²Includes 14:1n-7, 14:1n-5, 15:1n-5, 16:1n-5, 18:1n-5, 20:1n-9, and 20:1n-5.³Includes 22:5n-6 and 22:4n-6. ⁴Includes 16:3n-3 and 16:4n-3. ⁵LC- PUFA, long-chain polyunsaturated fatty acid (sum of 20:4n-3, 20:5n-3 22:5n-3 and 22:6n-3).

Table 3. Growth performance of meagre larvae fed the experimental feeds at 30 (initial) and 44 days post hatching (dph).

	Diets					
	Initial	C+	C-	EC	EECC	KRILL
Total length (mm)	8.8 ± 1.4	26.3 ± 4.2	25.6 ± 3.9	24.9 ± 4.3	26.0 ± 4.1	26.2 ± 4.3
Dry weight (mg)	1.1 ± 0.2	16.2 ± 4.0	16.6 ± 1.8	16.4 ± 4.7	19.2 ± 3.5	17.3 ± 3.7
SGR (% d ⁻¹)	-	16.8 ± 4.0	17.5 ± 1.8	16.9 ± 4.7	18.5 ± 3.5	17.2 ± 3.7
Survival (%)	-	19.5 ± 1.3	20.1 ± 1.2	20.9 ± 2.3	20.3 ± 1.0	19.7 ± 0.9

Data are means ± SD. dph, days post hatching; SGR, specific growth rate.

Table 4. Proximate composition and TBARs content in meagre larvae (44 dph) fed with the experimental diets.

Diets	C+			C-			EC		EECC		KRILL	
Proximate composition (%)												
Protein	11.0	±	1.1	10.4	±	2.1	10.6	±	1.4	10.9	±	2.0
Lipid	2.8	±	0.4	2.5	±	0.2	2.3	±	0.3	2.4	±	0.1
Moisture	82.9	±	2.8	83.9	±	2.1	83.7	±	2.1	82.7	±	2.2
Ash	2.5	±	0.6	2.8	±	0.1	2.9	±	0.2	3.1	±	0.1
TBARs content (μmol g ⁻¹ dry mass)	769.2	±	110.5 ^a	1028.8	±	159.3 ^a	862.2	±	136.3 ^a	138.5	±	45.7 ^b
	974.6	±	118.9 ^a									

Data expressed as means of three technical replicates per batch of larvae (n = 3). Different superscript letters denote differences among treatments identified by one-way ANOVA (P<0.05).

Table 5. Fatty acid composition (percentage of fatty acids) of meagre larvae fed with experimental diets at the end of the dietary trial (44 days post hatching).

<i>Fatty acids (%)</i>	C+			C-			EC			EECC			KRILL		
14:0	2.6	±	0.1	2.6	±	0.2	2.5	±	0.2	2.5	±	0.2	2.9	±	0.2
16:0	20.9	±	0.3 ^a	20.9	±	0.3 ^a	21.3	±	0.6 ^a	21.0	±	0.2 ^a	22.5	±	0.2 ^b
18:0	4.8	±	0.3	4.8	±	0.2	5.0	±	0.2	5.1	±	0.6	5.1	±	0.2
20:0	0.1	±	0.0 ^{ab}	0.2	±	0.0 ^b	0.1	±	0.0 ^a	0.0	±	0.0 ^{ab}	0.1	±	0.0 ^a
Σ Saturated¹	28.8	±	0.5 ^a	28.9	±	0.2 ^a	29.3	±	0.7 ^a	29.1	±	0.3 ^a	31.2	±	0.3 ^b
16:1n-7	3.6	±	0.1	3.6	±	0.2	3.4	±	0.1	3.4	±	0.1	3.5	±	0.3
18:1n-9	15.2	±	0.1 ^b	15.5	±	0.3 ^b	15.1	±	1.0 ^b	14.7	±	0.2 ^b	11.0	±	0.7 ^a
18:1n-7	5.3	±	0.1 ^a	5.5	±	0.0 ^a	5.4	±	0.0 ^a	5.3	±	0.1 ^a	5.9	±	0.1 ^b
20:1n-7	1.0	±	0.0 ^b	1.1	±	0.0 ^b	1.0	±	0.1 ^b	1.0	±	0.0 ^b	0.7	±	0.0 ^a
22:1n-11	0.3	±	0.0 ^b	0.3	±	0.0 ^b	0.3	±	0.1 ^b	0.2	±	0.1 ^b	0.0	±	0.0 ^a
Σ Monosaturated²	26.6	±	0.31 ^b	27.1	±	0.6 ^b	26.6	±	1.5 ^b	26.0	±	0.3 ^b	22.8	±	1.1 ^a
18:2n-6	6.6	±	0.1 ^b	6.5	±	0.2 ^b	6.6	±	0.0 ^b	6.6	±	0.1 ^b	4.5	±	0.1 ^a
18:3n-6	0.1	±	0.0 ^b	0.1	±	0.0 ^b	0.1	±	0.0 ^b	0.1	±	0.0 ^b	0.1	±	0.0 ^a
20:2n-6	0.2	±	0.0 ^b	0.2	±	0.0 ^b	0.2	±	0.0 ^b	0.2	±	0.0 ^b	0.1	±	0.0 ^a
20:3n-6	0.1	±	0.0 ^b	0.1	±	0.0 ^b	0.1	±	0.0 ^b	0.1	±	0.0 ^b	0.1	±	0.0 ^a
20:4n-6	0.9	±	0.2	0.9	±	0.1	0.9	±	0.1	1.0	±	0.3	0.9	±	0.1
Σ n-6 PUFA³	8.2	±	0.2 ^b	8.0	±	0.1 ^b	8.1	±	0.1 ^b	8.3	±	0.4 ^b	5.8	±	0.2 ^a
18:3n-3	1.4	±	0.0 ^b	1.4	±	0.0 ^b	1.3	±	0.1 ^b	1.4	±	0.2 ^b	0.9	±	0.0 ^a
18:4n-3	1.4	±	0.1 ^a	1.4	±	0.0 ^a	1.3	±	0.0 ^a	1.3	±	0.1 ^a	1.6	±	0.1 ^b
20:3n-3	0.1	±	0.0 ^b	0.1	±	0.0 ^b	0.1	±	0.0 ^b	0.1	±	0.0 ^b	0.1	±	0.0 ^a
20:4n-3	0.3	±	0.0 ^b	0.3	±	0.0 ^b	0.3	±	0.0 ^b	0.3	±	0.0 ^b	0.3	±	0.0 ^a
20:5n-3	13.7	±	0.8 ^a	13.7	±	0.3 ^a	13.7	±	0.6 ^a	13.7	±	1.2 ^a	16.6	±	1.4 ^b
22:5n-3	1.3	±	0.0	1.3	±	0.1	1.3	±	0.1	1.4	±	0.1	1.2	±	0.1
22:6n-3	16.0	±	0.6 ^a	15.9	±	0.8 ^a	16.0	±	0.6 ^a	16.3	±	0.4 ^a	17.6	±	0.2 ^b
Σ n-3PUFA⁴	34.4	±	0.4 ^a	34.1	±	0.9 ^a	34.1	±	1.1 ^a	34.6	±	0.8 ^a	38.3	±	1.5 ^b
Σ n-9PUFA⁵	15.9	±	0.1 ^b	16.3	±	0.3 ^b	15.9	±	1.1 ^b	15.5	±	0.2 ^b	11.8	±	0.6 ^a
(n-3+n-6) PUFA	8.2	±	0.2 ^b	8.0	±	0.1 ^b	8.1	±	0.1 ^b	8.3	±	0.4 ^b	5.8	±	0.2 ^a
Total n-3 LC-PUFA⁶	31.5	±	0.3 ^a	31.2	±	1.0 ^a	31.3	±	1.2 ^a	32.5	±	2.1 ^a	35.7	±	1.5 ^b
PIn	246.3	±	2.1 ^a	243.7	±	7.7 ^a	244.4	±	8.2 ^a	248.5	±	3.9 ^a	275.3	±	3.0 ^b

Data expressed as means of three technical replicates per batch of larvae (n = 3). Different superscript letters denote differences among treatments identified by one-way ANOVA (P<0.05). ¹Includes 15:0 and 17:0. ²Includes 14:1n-7, 14:1n-5, 15:1n-5, 16:1n-5, 18:1n-5, 20:1n-9 and 20:1n-5. ³Includes 22:5n-6 and 22:4n-6. ⁴Includes 16:3n-3 and 16:4n-3. ⁵Includes. 22:1n-9, 20:3n-9, 20:2n-9, 20:1n-9, 18:2n-9, 18:1n-9. ⁶ LC-PUFA, long-chain polyunsaturated fatty acid (sum of 20:4n-3, 20:5n-3, 22:5n-3 and 22:6n-3). PIn, peroxidation index.

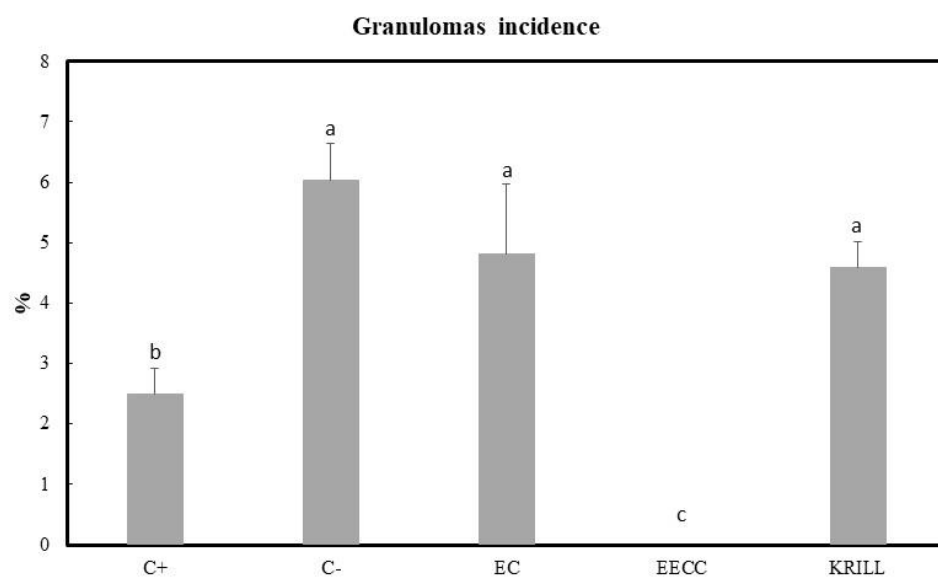


Figure 1

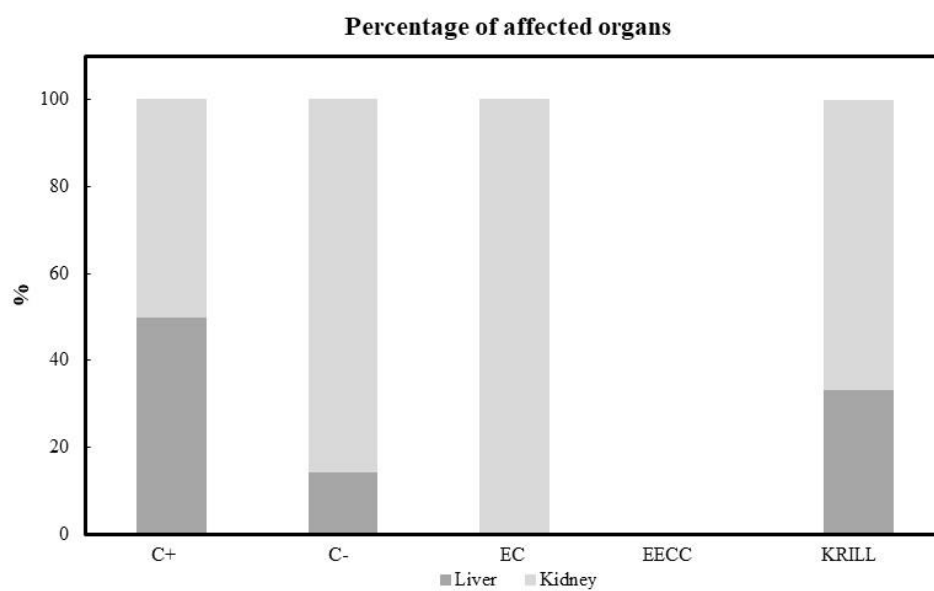


Figure 2

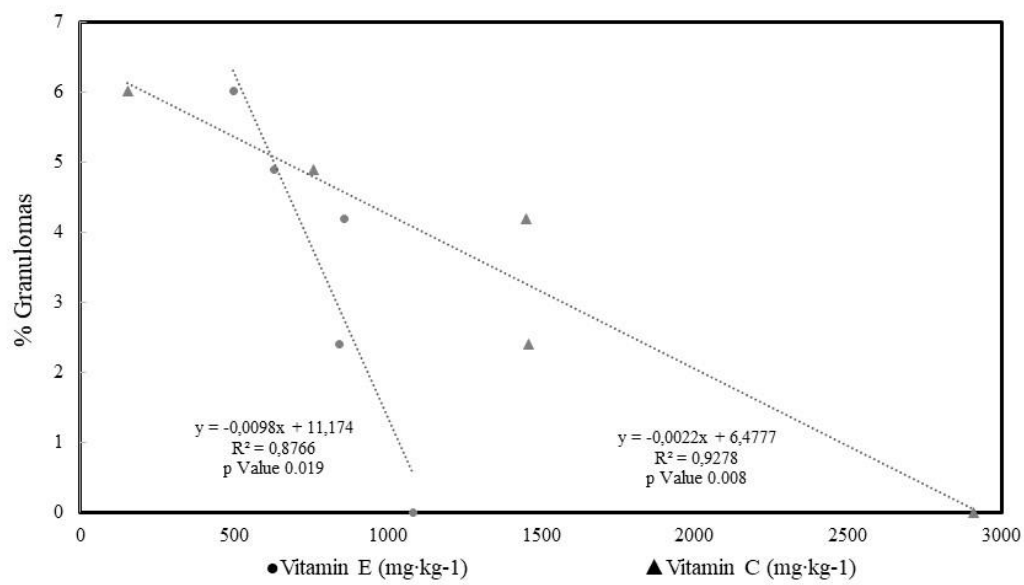
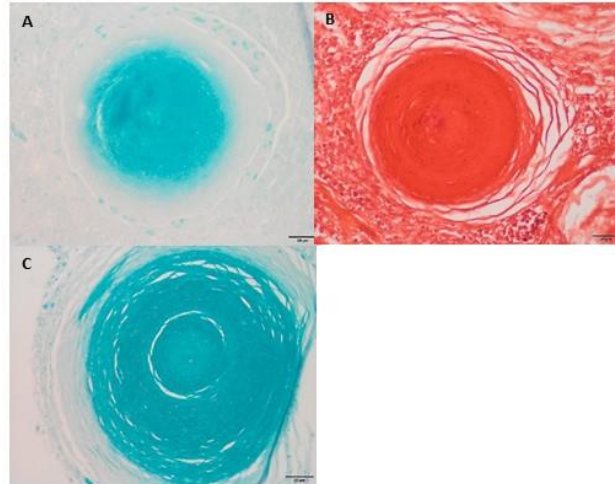
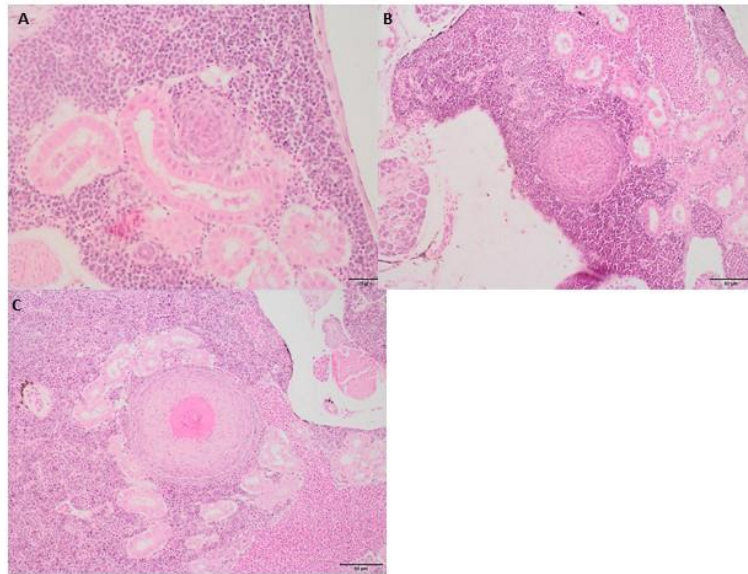


Figure 3



Supplementary Figure 1



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Supplementary Figure 2